

A NEW FLAVONE GLUCOSIDE FROM *Stachys aegyptiaca*

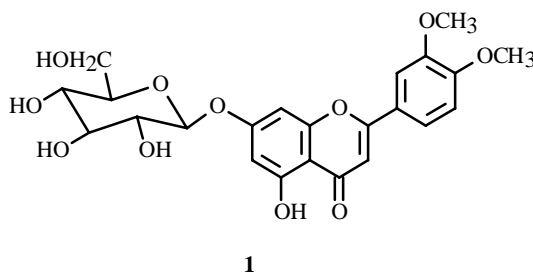
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In addition to 5,7,3'-trihydroxy-6,4'-dimethoxyflavone, 5,7,3'-trihydroxy-6,8,4'-trimethoxyflavone, isoscutellarein, luteolin, and luteolin-7-O-glucoside, the methanol extract of the aerial parts of *Stachys aegyptiaca* yielded a new flavone identified as luteolin 3',4'-dimethylether-7-O- β -D-glucoside on the basis of chemical and spectroscopic methods.

Key words: *Stachys aegyptiaca*, Lamiaceae, new flavone glucoside.

Stachys aegyptiaca is a perennial plant growing wild in the Sinai desert [1] and proved to be a rich source for flavonoids. We have previously reported the isolation and structure elucidation of diapigenin 7-O-(6''-trans-6''-cis-p,p'-dihydroxy- μ -truxinyl)glucoside [stachysetin] [2] and hypoluetin 7-[6''-acetylallosyl-(1 \rightarrow 2)-3''-acetyl-glucoside, along with 24 known flavonoid aglycones and glycosides [3]. Also, the acetylated isoscutellarein glucoside was reported [4]. In continuation of our study on the phenolic constituents of *S. aegyptiaca*, the present communication describes the further isolation and structure elucidation of a new flavone glycoside, the structure of which was identified as luteolin 3',4'-dimethylether-7-O-glucoside from the aerial parts of the same plant.



The methanol extract of the aerial parts of *S. aegyptiaca* was fractionated on a polyamide column with H₂O as eluent with increasing amounts of MeOH. Isolation and purification were achieved by combination of PPC, TLC on silica gel and Sephadex LH-20.

Flavonoid **1** was isolated as a yellowish white amorphous powder. Acid hydrolysis of **1** afforded glucose, which was identified by co-chromatography with authentic samples and the aglycone **1a**. The UV spectral data of **1a** with diagnostic shift reagents [5] indicated a flavone, absence of a free *ortho*-dihydroxyl pattern, occupation at positions-3' and -4', and a free hydroxyl group at position-7. The UV spectral data of **1** indicated a flavone with occupation at positions-7,3' and -4' [5].

The EI-MS spectrum of permethylated [6] **1** gave a molecular ion peak at m/z 546 (66%) in accordance with fully methylated luteolin glucoside. A fragment at m/z 328 was due to the aglycone luteolin bearing three methoxyl groups. The identity of the glucose moiety was confirmed by the fragment at m/z 218 due to the ion [glucose-H]⁺ and the fragment at m/z 187 for the ion [glucose-MeOH]⁺.

The ¹H NMR spectrum of **1** showed that it is a monoglucoside of luteolin dimethylether on the basis of the signal H-1 glucose (δ 5.1), and the coupling constant ($J = 7.5$ Hz) is characterized for a β -linked glucose. Furthermore, the chemical shift (δ 5.1) confirmed that the glucose is directly attached to the aglycone [7]. The spectrum also showed signals at δ 7.4 and δ 6.9 (m; d, $J = 8.0$ Hz) assigned for H-2',6' and H-5', confirming the disubstituted pattern at ring-B. Two doublets ($J = 2.5$ Hz) at

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δ 6.8 and δ 6.4 were assigned for H-8 and H-6, while H-3 appeared as a singlet at δ 6.75. The two methoxyls groups appeared as two singlets at δ 3.95 and δ 3.90.

The ^{13}C NMR spectrum confirmed that **1** is a monoglucoside of luteolin on the basis of C-6 glucose (δ 61.0). The ^{13}C NMR shifts of the aglycone part of **1** correspond with the shifts of luteolin [8], the only difference being a downfield shift of the signal assigned to C-3' and C-4' by approximately 3.7 ppm, confirming the location of the $-\text{OCH}_3$ groups at C-3' and C-4'. Also the signal assigned to C-7 was shifted upfield by approximately 2 ppm and there was a downfield shift of about 1.8 ppm for the *para*-related carbon (C-10). These shifts are analogue to those reported when the hydroxyl group is glycosylated at C-7 in flavonoids [9 – 11]. The assignments of the glucose carbons in **1** are based on those given in the literature [7, 11, 12]. From the above data, compound **1** is identified as luteolin 3',4'-dimethylether-7-*O*- β -D-glucoside.

EXPERIMENTAL

Plant Material. *S. aegyptiaca* aerial parts were collected around the Suez Canal University guest house at Saint Catherine, Sinai, in March 2004.

Extraction and Isolation. Dried plant (1.25 kg) was extracted with 80% MeOH. The concentrated extract was subjected to a polyamide column eluted with a H_2O -MeOH mixture with increasing amounts of MeOH. PPC using H_2O , 15% AcOH, BAW (*n*-BuOH-AcOH- H_2O , 4:1:5, upper phase) afforded pure samples of the known flavonoids and the aglycone **1a**. A combination of TLC (CHCl_3 -MeOH- H_2O , 45:30:2) and Sephadex LH-20 (MeOH) afforded compound **1**.

Luteolin 3',4'-dimethylether-7-*O*- β -D-glucoside (1). UV spectra (MeOH, λ_{max} , nm): 255, 267, 352; +NaOMe: 265, 397; + AlCl_3 : 273, 297, 330sh, 422; AlCl_3 +HCl: 260, 275, 297, 350, 385; NaOAc: 260, 370; NaOAc+ H_3BO_3 : 260, 370. ^1H NMR (270 MHz, $\text{DMSO}-d_6$, δ , J/Hz): 7.40 (2H, m, H-2',6'), 6.90 (1H, d, J = 8.0, H-5'), 6.80 (1H, d, J = 2.5, H-8), 6.75 (1H, s, H-3), 6.40 (1H, d, J = 2.5, H-6), 5.1 (1H, d, J = 7.5 Hz, H-1 glu), 3.95 (3H, s, $-\text{OCH}_3$), 3.90 (3H, s, $-\text{OCH}_3$). ^{13}C NMR (270 MHz, $\text{DMSO}-d_6$, δ): 55.0 ($-\text{OCH}_3$), 55.8 ($-\text{OCH}_3$), 61.0 (C-6''), 69.6 (C-4''), 73.5 (C-2''), 76.2 (C-3''), 77.2 (C-5), 94.5 (C-8), 99.8 (C-6), 100.0 (C-1), 103.0 (C-3), 105.0 (C-10), 111.0 (C-2'), 120.0 (C-6'), 124.5 (C-1'), 149.5 (C-3'), 151.5 (C-4'), 152.5 (C-5'), 158.9 (C-9), 161.0 (C-5), 162.7 (C-7), 164.4 (C-2), 182.0 (C-4).

REFERENCES

1. V. Tackholm, *Student's Flora of Egypt*, Cairo University, Cairo, 1974, p. 464.
2. M. A. El-Ansari, D. Barron, M. F. Abdalla, N. A. M. Saleh, and J. L. Le Quere, *Phytochemistry*, **30**, 1169 (1991).
3. M. A. El-Ansari, M. A. Nawwar, and N. A. M. Saleh, *Phytochemistry*, **40**, 1543 (1995).
4. M. Sharaf, *Fitoterapia*, **69** (4), 35 (1998).
5. T. J. Mabry, K. R. Markham, and M. B. Thomas, *The Systematic Identification of Flavonoids*, Springer, Berlin, 1970.
6. S. Hokomori, *J. Biochem.* (Tokyo), **55**, 205 (1964).
7. B. O. Osterdahl and G. Lindberg, *Acta Chem. Scand.*, **B31**, 293 (1979).
8. K. R. Markham and V. M. Chari, *The Flavonoids: Advances in Research*, In: J. B. Harbone, T. J. Mabry, editors, Chapman and Hall, London, 1982.
9. M. Sharaf, M. A. El-Ansari, and N. A. M. Saleh, *Fitoterapia*, **69**, 47 (1998).
10. K. R. Markham and B. Ternai, *Tetrahedron*, **32**, 2607 (1976).
11. K. R. Markham, B. Ternai, R. Stanley, H. Geiger, and T. J. Mabry, *Tetrahedron*, **34**, 1389 (1978).
12. O. B. Osterdahl, *Acta Chem. Scand.*, **B30**, 867 (1976).